

1 **The toxicity of potentially toxic elements (Cu, Fe, Mn, Zn and Ni) to**  
2 **the cnidarian *Hydra attenuata* at environmentally relevant**  
3 **concentrations**

4  
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14 **1. Introduction**

15 Pollution of the aquatic environment is common near human activity and the  
16 presence of chromium (Cr), copper (Cu), manganese (Mn), nickel (Ni), lead (Pb), tin  
17 (Sn) and zinc (Zn) is associated with antifouling paint, industry, insecticides, fertilizer  
18 use, fuel consumption and waste water treatment works (Caccia *et al.*, 2003; Deheyn  
19 and Latz, 2006; Canning-Clode *et al.*, 2011; Xu *et al.*, 2014; Rodriguez-Iruretagoiena  
20 *et al.*, 2016). Areas with intense, localised activity (e.g. harbours and ports within  
21 estuaries) are known to exhibit greater anthropogenic influence (Birch *et al.*, 2015).  
22 An example of this is the Clyde Estuary, Scotland, which has since the Industrial  
23 Revolution received pollution from ship building, dye works and petroleum  
24 installations resulting it becoming the UK's most contaminated estuarine environment  
25 (Turner, 2000; Edgar *et al.*, 2003; Vane *et al.*, 2007; Vane *et al.*, 2011).

26 The Water Framework Directive (2000/60/EC) and Annex II of the  
27 Environmental Quality Standards Directive (2008/105/EC) include a number of PTEs  
28 (arsenic (As), cadmium (Cd), mercury (Hg), nickel (Ni)) which are classified as  
29 'priority substances' and 'priority hazardous substances'. Requirements of the Water  
30 Framework Directive include a target of 'good' ecological status for waterbodies and

31 levels of pollutants within the Clyde have reduced over the last three decades and  
32 ecological recovery has been observed (Critchlow-Watton *et al.*, 2014).

33 Elements such as Cu, Fe, Zn and Mn, are essential in trace amounts to  
34 support and maintain functions in aquatic ecosystems (Tchounwou *et al.*, 2012),  
35 however Pb, Cu and Zn have previously found to be a 'triad' of metals associated  
36 with human influence, and at high concentrations are toxic, adversely impacting on  
37 human/animal health and the environment (ten Brink and Woudstra, 1991; McLellan  
38 *et al.*, 2013). To determine the effect of pollutants on aquatic organic substances,  
39 numerous studies exposing invertebrates to heavy metals have previously been  
40 undertaken (Lasier *et al.*, 2000; Borgmann *et al.*, 2005; Torres Guzmán *et al.*, 2010;  
41 García *et al.*, 2011; Liber *et al.*, 2011; Lopes *et al.*, 2014, Joško *et al.*, 2016).  
42 Karntanut and Pascoe (2002) exposed four different *Hydra* species (*vulgaris*  
43 (*Zurich*), *attenuata*, *oligactis* and *viridissima*) to varying concentrations of Cu, Cd and  
44 Zn. A variation in the lethal concentration (LC) between species was found with Cu  
45 being the most toxic of the three elements with an LC<sub>50</sub> ranging from 0.025 to 0.084  
46 mg/l after 96h of exposure, followed by Cd (0.16 to 0.52 mg/l), then Zn (11 to 14  
47 mg/l).

48 *H. attenuata* (also known as *Hydra vulgaris*) is a species of cnidarian that are  
49 ubiquitously found in freshwater ecosystems and are commonly used for toxicity  
50 testing. The health status and acute toxicity of *H. attenuata* is easily observed  
51 through a series of defined morphological changes following exposure to a toxin in a  
52 relatively simple bioassay (Wilby, 1988). Other chronic endpoints used to measure  
53 toxicity include asexual reproduction (budding), feeding behaviour and attachment to  
54 a substrate (Quinn *et al.*, 2012). This species has relatively unique regenerative  
55 properties, is easy to culture and maintain in a laboratory, has a high reproductive

56 rate and as a diploblastic organism, is sensitive to environmental pollutants and is  
57 therefore used as a bioindicator for the health of a freshwater aquatic ecosystem  
58 (Quinn *et al.*, 2012). *H. attenuata* have been widely used in cost effective bioassays  
59 to assess the toxicity of numerous contaminants including wastewater (Trottier *et al.*,  
60 1997), industrial effluents (Blaise and Kusui, 1997), pharmaceuticals (Pascoe *et al.*,  
61 2003; Quinn *et al.*, 2008a, 2008b, 2009) , PTEs (Holdway *et al.*, 2001; Karntanut and  
62 Pascoe, 2002; Quinn *et al.*, 2007) and more recently rare earth elements (Blaise *et*  
63 *al.*, 2018).

64 The aim of this study is to evaluate the toxicity of PTE's, both individually and  
65 as a mixture, found in the Clyde estuary in Scotland against the cnidarian *Hydra*  
66 *attenuata*. Water samples from various locations in the Clyde estuary (Scotland)  
67 were analysed for anthropogenic PTEs Cu, Fe, Mn, Ni and Zn. The toxicity of these  
68 metals individually and as a mixture at environmentally relevant and elevated  
69 concentrations was tested using the *H. attenuata* bioassay on the ecologically  
70 relevant endpoints of morphology, feeding, attachment and reproduction. To the best  
71 of our knowledge, this is the first toxicity study investigating the mortality of any  
72 *Hydra* species individually exposed to Fe, Mn or Ni.

73

## 74 **2. Materials and Methods**

### 75 **2.1. Test organism**

76 *Hydra* were maintained in glass bowls containing 0.5 L of *Hydra* media (147 mg/l  
77 CaCl<sub>2</sub>H<sub>2</sub>O, 110 mg/l TES [N-Tris(hydroxymethyl) methyl 1-2-aminoethanesulfonic  
78 acid], pH adjusted to 7 using 0.5 M NaOH), maintained at 18± 2°C with a 12 h light  
79 12 h dark photoperiod. *Hydra* were fed 3 times per week with newly hatched *Artemia*

80 *salina* nauplii and were fasted 48 h prior to exposure. To avoid algal contamination,  
81 *Hydra* media was regularly changed after each feeding.

82

## 83 **2.2. PTE determination**

84 Water samples were collected from two estuarine (Kelburn Park and Erskine  
85 Harbour) and one freshwater (Gourock Burn) sites along the Clyde estuary (Fig. 1).  
86 Samples were collected in polypropylene sample bottles with pH and temperature  
87 recorded immediately (Mettler Toledo). Samples were then acidified with conc.  
88 HNO<sub>3</sub> (Fisher Trace Grade, UK) on site for preservation, transported to the laboratory  
89 and refrigerated at 4°C until analysis. Prior to analysis samples were filtered to  
90 <45µm (Filtermate, Environmental Express, USA).

91 Potentially toxic elements were determined by ICP-OES (Thermo Fisher,  
92 iCAP); a calibration series (0 mg/l, 2 mg/l and 10 mg/l of multi-element standard,  
93 ME/1001/05; Fisher Scientific, UK) was determined. Samples were analysed in  
94 triplicate. ICP-OES conditions were as follows: rf generator: 1.15 kW; Plasma: 1.4  
95 l/min; Auxillary: 0.5 l/min; Nebuliser: 0.8 l/min; sample flow rate 1.5 ml/min.

96 Averages of the sample concentrations were calculated; Limits of Detection were  
97 calculated using standard practice (e.g. (McLellan *et al.*, 2013)) (Table 1). It can be  
98 seen that levels of Fe within the Gourock Burn were very high therefore it was  
99 decided not to put this forward at the reported concentrations. Ni was taken at 0.5  
100 mg/l to reflect potential toxicity levels within the selected biota. These are the  
101 'environmentally relevant' concentrations.

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103

104 **2.3. Test solutions**

105 Environmental relevant PTE solutions (1x) of metals in hydra media (HM) were  
106 prepared for the individual elements and for the mix solution. These stock solutions  
107 were then diluted with HM to give concentrations: 0.0001x, 0.001x, 0.01x, 0.1x, 10x,  
108 100x and 1000x concentrations were made from a 1000x stock solution (Table 2).

109

110 **2.4. Hydra toxicity tests**

111 All PTE exposures to *Hydra attenuata* were undertaken in quadruplicate (4  
112 repetitions of each concentration) and the whole experiment was undertaken in  
113 triplicate for the PTE mixture and duplicate for each individual metal exposure. A 4 ml  
114 sample of the relevant solution was added to 4 wells in a 24 multiwell plate,  
115 containing a single *Hydra*, and the wells wrapped in parafilm to prevent evaporation  
116 and kept at  $18 \pm 2^\circ\text{C}$  for 96 h. Healthy *Hydra* with a morphology score of 10 on the  
117 Wilby table (Table 3) and having one bud (2 hydranths) were used in each exposure.  
118 Selection of healthy *Hydra* was undertaken using a binocular microscope.  
119 Morphology, hydranth number and attachment were observed at 24, 48, 72 and 96 h.  
120 The *Hydra's* ability to ingest prey (feeding endpoint) was tested on all *Hydra* that  
121 scored  $> 5$  on the Wilby score table after 96 h as per Quinn et al., (2007). These  
122 *Hydra* were placed individually into a well of a clean 24 well multi-well plate  
123 containing 4 ml of *Hydra* media. Freshly hatched *Artemia* were rinsed three times  
124 with HM with 5 individuals added to each well at time 0, taking care not to add them  
125 directly to the tentacles of the *Hydra*. The number of ingested prey were observed  
126 every 20 min for 120 min.

127

## 128 **2.5. Statistical Data Analysis**

129 The 96 h LC<sub>50</sub> values for the mortality exposure were calculated using the Probit  
130 analysis program. The mortality exposure the sub-lethal LOEC (Lowest Observable  
131 Effect Concentration) was reported for  $\geq 2$  *Hydra* with score 8 or below and the NOEC  
132 (No Observable Effect Concentration) was based on *Hydra* with a score  $>8$  (Quinn et  
133 al., 2009). A toxicity threshold (TT) was determined from the LOEC and NOEC using  
134 the following equation:  $TT = (NOEC \times LOEC) / 2$  (US EPA, 1989). Variability in all  
135 endpoints (morphology, attachment, hydranth number, feeding behaviour) between  
136 the exposed and control *Hydra* were tested by one-way analysis of variance  
137 (ANOVA). Significance was set at  $p \leq 0.05$ . The Pearson correlation coefficient was  
138 used to measure the strength of the association between the concentration of the  
139 pollutant and the endpoints.

140

141

## 142 **3 Results**

### 143 **3.1. Toxicity of individual metals to *Hydra attenuata***

144 Complete (100%) population mortality (indicated by a score  $\leq 5$  on the Wilby scale)  
145 was found at 0.1x (0.05 mg/l) for Cu (Fig. 2 A), 0.1x (0.3 mg/l) for Fe (Fig. 2 C), 10x  
146 (5 mg/l) for Ni (Fig. 2 D). For Mn, 100% mortality of all *Hydra* exposed was found at  
147 100x (200 mg/l). Although some mortality was observed at 1x, mortality numbers  
148 were low (Fig. 2 B). Highly significant ( $p = < 0.005$ ) and negative correlations were  
149 found with hydranth number and feeding behaviour (Table 5). For Zn, 100% mortality  
150 was found at 1000x (100 mg/l). Although mortality was detected at 100x (10 mg/l),  
151 mortality numbers were low (Fig. 2 E). An extremely significant ( $p = < 0.001$ ) and

152 negative correlation was found with hydranth number, and a very significant ( $p = <$   
153 0.005) negative correlation was found for feeding behaviour (Table 5). The 96 h LC<sub>50</sub>  
154 values were determined as follows: Cu 0.0225 mg/l, Mn 20 mg/l, Fe 0.135 mg/l, Ni  
155 2.25 mg/l, and Zn 31.622 mg/l. The Toxicity thresholds were calculated at: Cu  
156 0.000125 mg/l, Mn 0.2 mg/l, Fe 0.000045 mg/l, Ni 0.0125 mg/l, Zn 5 mg/l (Table 4).

157

### 158 **3.2 Toxicity of PTE mixture**

159 For the PTE mixture, 100% mortality was found at 0.1x (Fig. 3). The 96h LC<sub>50</sub> value  
160 was calculated as 0.045x. The LOEC was 0.01x and NOEC was 0.001x. The toxicity  
161 threshold was calculated at 0.000005x (Table 4). The high toxicity of Cu was not  
162 entirely responsible for the very high toxicity of the mixture. The toxicity threshold for  
163 the mixture (0.000005x) showed that the mixture was more toxic than Cu individually,  
164 which had a toxicity threshold of 0.000125 mg/l (0.00025x).

165

### 166 **3.3 Toxicity of heavy metals at environmental concentration**

167 Both Cu and Fe when exposed individually to the concentration of their respective  
168 metals found in the environment resulted in 100% mortality of all *Hydra* exposed (Fig.  
169 2 A & C). A significant ( $p = < 0.001$ ) toxic effect occurred when *Hydra* were exposed  
170 to Mn and Ni at the environmentally relevant concentration. Zn remained at a perfect  
171 morphology score of 10 when exposed to the Zn concentration found in the  
172 environment (Fig. 2 E). The concentration of Zn found in the environment also had no  
173 significant toxic effect on hydranth number, feeding behaviour or attachment of *Hydra*  
174 to a substrate. When exposed to the concentration of Mn found in the environment, a  
175 significant ( $p = < 0.01$ ) toxic effect occurred in the attachment of *Hydra* to a substrate  
176 (Fig. 2 B). The concentration of Mn found in the environment had no significant toxic

177 effect on hydranth number or feeding behaviour. The concentration of Ni found in the  
178 environment resulted in a significant ( $p = < 0.05$ ) toxic effect on the feeding behaviour  
179 of *Hydra* (Fig. 2 D). The concentration of Ni found in the environment had no  
180 significant toxic effect on hydranth number or attachment of *Hydra* to a substrate.  
181 *Hydra* morphology was monitored at 24 h, 48 h, 72 h and 96 h of exposure to the  
182 concentration found in the environment with any toxic effect occurring within the first  
183 24 h of exposure (Fig. 4).

184

#### 185 **4 Discussion**

186 Since the 18<sup>th</sup> Century and the beginning of the Industrial Revolution, the Clyde has  
187 had a diverse heritage and there is a well-documented legacy of pollutants e.g.  
188 (Hursthouse *et al.*, 1994; Edgar *et al.*, 2003; Vane *et al.*, 2007; Vane *et al.*, 2011).  
189 The sample locations chosen for this site are near former landfill sites (Gourock Burn  
190 and Kelburn Park) or wastewater treatment works (Erskine Harbour) and there is  
191 potential for continued contamination from these sources. This is in addition to former  
192 industrial activity e.g. metal plating near Kelburn Park (Miller, 1986). Despite the  
193 improving physical and ecological status of the outer Clyde estuary (Critchlow-  
194 Watton *et al.*, 2014), it is concerning that this study has found that PTE levels are  
195 above legislative requirements (Table 6) which may be caused by the proximity of  
196 potential point sources of pollutants. In that light, the Clyde is similar to other  
197 estuaries where point sources can be attributed to elevated PTE levels (Larrose *et*  
198 *al.*, 2010; Birch *et al.*, 2015; Petit Jérôme *et al.*, 2015; Rodriguez-Iruretagoiena *et al.*,  
199 2016). Levels of all heavy metals tested were higher than levels in the Thames river  
200 in London, Canada (Environment and Engineering Services, 2018) and the Ganga  
201 river in India (Central Water Commission, 2018) (Table 6). The maximum acceptable



202 limits for copper (0.00376 mg/l) and iron (1 mg/l) based on EU / UK legislative  
203 requirements are higher than the *H. attenuata* LC<sub>50</sub>'s for copper (0.0225 mg/l) and  
204 iron (0.135 mg/l).

205 To the best of our knowledge, this is the first toxicity study investigating the  
206 toxicity of any *Hydra* species exposed to Fe, Mn or Ni. This study calculated the LC<sub>50</sub>  
207 values, LOEC, NOEC and Toxicity Thresholds for Cu, Fe, Mn, Zn and Ni (Table 4).  
208 The 96 h LC<sub>50</sub> results for Cu (0.0225mg/l) are similar to those reported by Karntanut  
209 and Pascoe (2000) (0.032 mg/l) for *H. vulgaris* (also known as *H. attenuata*) and for 4  
210 different species of Hydra; *H. vulgaris* Zurich (0.042 mg/l), *H. vulgaris* (0.056 mg/l), *H.*  
211 *oligactis* (0.084 mg/l), *H. viridissima* (0.025 mg/l) (Karntanut and Pascoe, 2002). The  
212 Cu LC<sub>50</sub> value in the current study were higher than the LOEC value which is unusual  
213 but is due to the dilution range used for the serial dilution.

214 The 96 h LC<sub>50</sub> value calculated for Zn in the present study (31.6 mg/l) is higher  
215 than those reported for *H. vulgaris* (7.4 mg/l) (Karntanut & Pascoe, 2000) *H. vulgaris*  
216 *Zurich* (14 mg/l), *H. vulgaris* (13 mg/l), *H. oligactis* (14 mg/l), *H. viridissima* (11 mg/l)  
217 (Karntanut & Pascoe, (2002). In the current study Zn was tested at a concentration of  
218 10 mg/l and a mortality percentage of 12.5% was found. The large divisions used in  
219 the serial dilutions resulted the high LC<sub>50</sub> value of 30 mg/l that was calculated, as the  
220 next concentration tested after 10 mg/l was 100 mg/l.

221 The same could be true of Mn (with an LC<sub>50</sub> value of 20 mg/l) but as this is the  
222 first time this metal has been used in a toxicity test to study mortality of *Hydra*, there  
223 is no literature for comparison. Harford *et al.*, (2015) however, exposed *Hydra*  
224 *viridissima* to varying levels of Mn to test population growth. The highest  
225 concentration tested by Harford *et al.*, (2015) was 10 mg/l at which the population of

226 *H. viridissima* was still growing but had dropped to 10% growth compared to the  
227 control. A very significant ( $p = <0.005$ ) negative correlation was found for hydranth  
228 number and feeding behavior when exposed to Mn.

229 An extremely significant ( $p = <0.001$ ) negative correlation for hydranth number  
230 and a very significant ( $p = <0.005$ ) negative correlation was found when exposed to  
231 Zn. However, there was no significant correlation for attachment when exposed to  
232 any of the tested metals and no significant correlation for attachment, hydranth  
233 number or feeding behavior when exposed to Cu, Fe or Ni.

234 In this study, a significant toxic effect occurred when *Hydra* were exposed to  
235 the Cu, Fe, Mn and Ni at concentrations found in the Clyde estuary (Fig. 2A-D).  
236 *Hydra* morphology was unaffected and remained at a score of 10 when exposed to  
237 the concentration of Zn found in the environment (Fig. 2E). Mortality levels of 100%  
238 were measured when *Hydra* were exposed to the heavy metal mixture (Fig. 3) and to  
239 Cu and Fe (individually) (Fig. 2A, C) at concentrations found in the Clyde. These  
240 results indicate that *Hydra attenuata* are unable to survive in aquatic environments  
241 with the metal concentrations found in the Clyde estuary, which may also have an  
242 impact on *Hydra* predators and prey.

243 The results also indicate that the PTE mixture (including the individual  
244 concentrations of Cu, Fe, Mn and Ni) could potentially prove significantly toxic to  
245 other invertebrates. The concentration of Cu found in the Clyde estuary was  
246 measured at 0.5 mg/l, this was 22 times higher than the LC<sub>50</sub> found for *Hydra*  
247 *attenuata*. When compared with other studies (Table 7), the levels of Cu found in the  
248 Clyde would also be toxic to aquatic vertebrates such as *Rasbora sumatrna*, the  
249 guppy (*Poecilia reticulata*) and the zebrafish (*Danio rerio*). The concentration of Fe

250 found in the Clyde estuary was measured at 3 mg/l, this was also 22 times higher  
251 than the LC<sub>50</sub> found for *Hydra attenuata* and would be toxic to other aquatic  
252 invertebrates such as *Daphnia magna*, and aquatic vertebrates, such as the brown  
253 trout (*Salmo trutta*) (Table 7).

254 For the PTE mixture, a significant ( $p \leq 0.05$ ) toxic effect was seen at the lowest  
255 concentration studied (0.0001x) (Fig. 3). The LC<sub>50</sub> was calculated as 0.045x for the  
256 mixture and the toxicity threshold was calculated as 0.000005x. The toxicity threshold  
257 (TT) was lower than any of the corresponding values of the individual metals  
258 contained within the mixture (Table 4). This result indicates that the metals have a  
259 cumulative effect, with each metal behaving cumulatively, contributing to the total  
260 effect of the mixture and further increasing the toxicity.

261 Morphology was found to be the most significant endpoint in studying the toxic  
262 effects of metals. Using the additional endpoints of hydranth number, attachment and  
263 feeding behavior, Quinn *et al.*, (2007) found a significant decrease in hydranth  
264 number, attachment and feeding behavior as the concentration of the toxin  
265 increased. In the present study, a similar significant negative correlation was  
266 observed for hydranth number and feeding behavior following exposure to Mn and  
267 Zn. There was no significant correlation found with attachment in any of the  
268 exposures undertaken.

269 Most toxicity tests involving *Hydra* spp expose the organism to a toxin for 96 h.  
270 In this study, it was observed that any significant toxic effect of a pollutant occurred  
271 within the first 24 h of exposure. A review of other toxicity studies using *Hydra* as a  
272 test organism shows that the toxic effect of a contaminant occurs within the first 24 h  
273 of exposure (Blaise and Kusui, 1997; Karntanut and Pascoe, 2000, 2002). It may

274 therefore be necessary to only expose *Hydra* to a toxin for 24 h to test a compounds  
275 toxicity. However, more research is needed to confirm this. The potential  
276 replacement of a 96 h exposure with a 24 h one would greatly reduce the time  
277 needed for toxicity testing, helping to reduce the cost and potentially increasing the  
278 number of toxins that can be tested within a given time period.

279

## 280 **5. Conclusion**

281 This paper shows that a significant toxic effect was observed on *Hydra* exposed to  
282 the PTE mixture at the concentration found in the environment (1x) after a short-term  
283 exposure period (24 h). The high toxicity of Cu was not entirely responsible for the  
284 very high toxicity of the mixture. The toxicity threshold for the mixture (0.000005x)  
285 showed that the mixture was more toxic than Cu individually, which had a toxicity  
286 threshold of 0.000125 mg/l (0.00025x). The toxicity threshold (TT) for the PTE  
287 mixture was lower than that for the same metals when exposed individually to *Hydra*,  
288 indicating that metals may act cumulatively in a mixture. However, a significant toxic  
289 effect occurred when *Hydra* were exposed individually to Cu, Fe, Mn and Ni at  
290 concentrations found in the environment, with 100% mortality when exposed  
291 individually to the environmental concentrations of Cu and Fe. These high  
292 environmental concentrations of PTE would impact, not only on the predator and  
293 prey interactions within the *Hydra* community but also could potentially prove  
294 significantly toxic to other aquatic organisms.

295

296

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300

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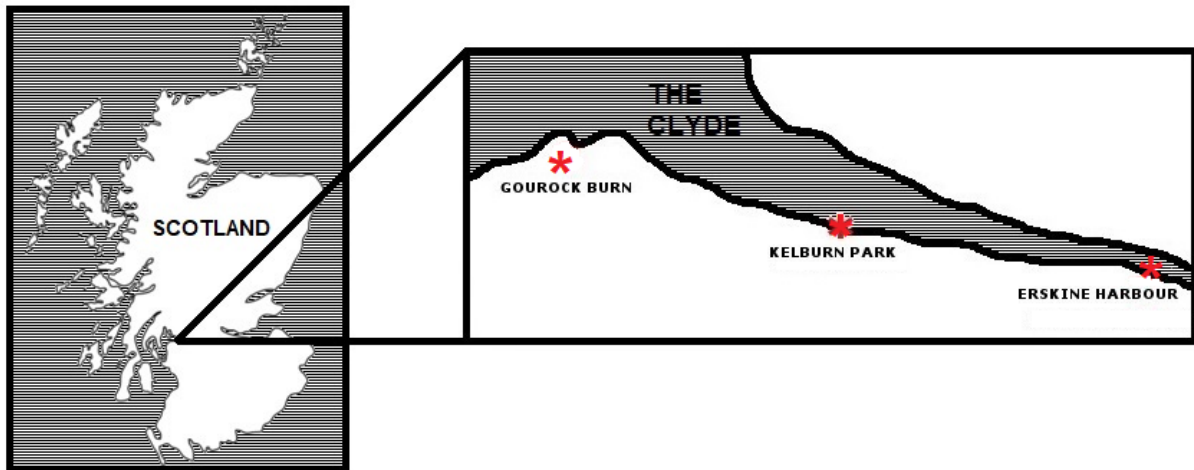
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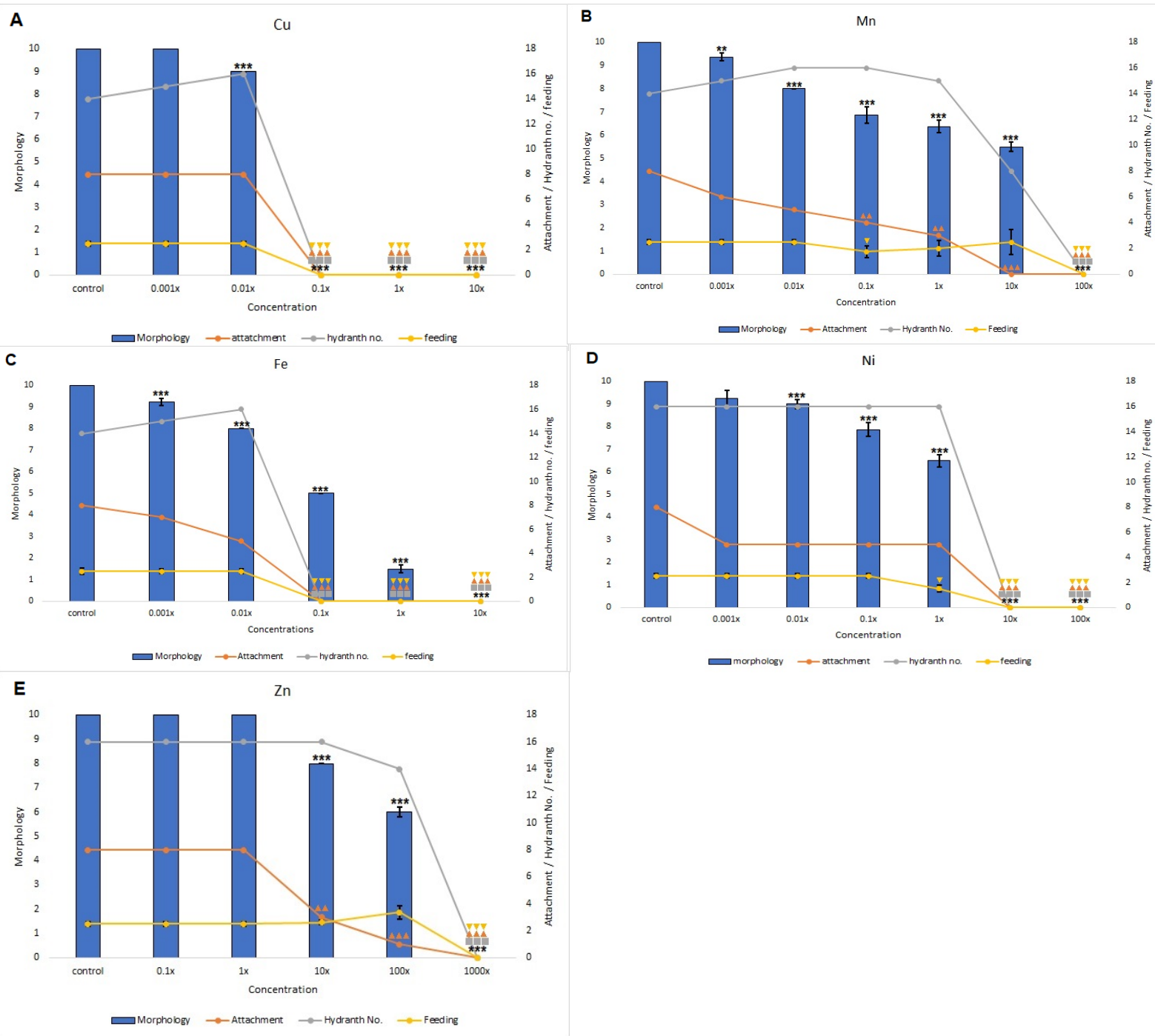


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457 Fig. 1: Overview of heavy metal sampling locations along the Clyde estuary, Scotland. \* indicates  
458 sample location.

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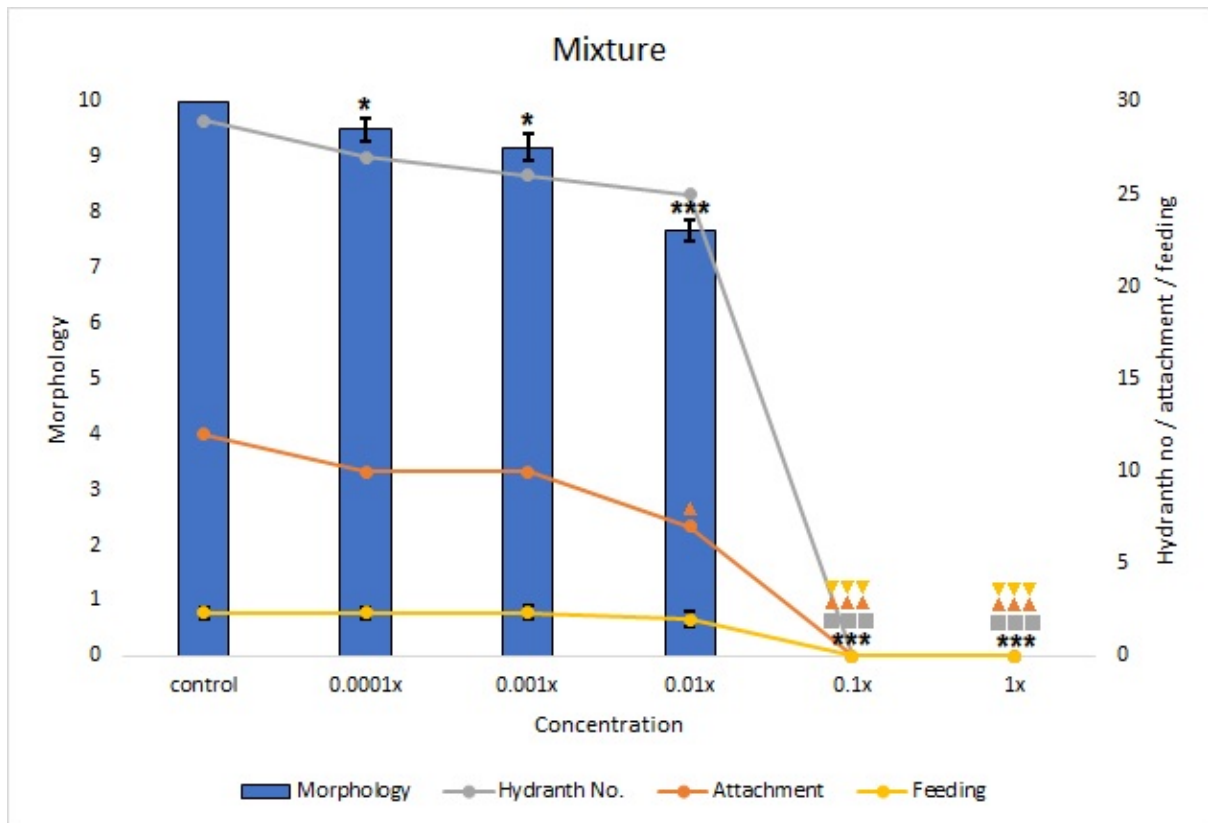
462 Fig. 2: – Lethal and sub-lethal effects of individual metals at varying concentrations on *Hydra*  
 463 morphology, hydranth number, attachment and feeding after 96hr exposure. Points at Morphology and  
 464 feeding represent the mean score (n=8)±standard error. Points at attachment and hydranth number  
 465 represent sum(n=8). Significance for morphology at \*= $p \leq 0.05$ ; \*\*= $p \leq 0.01$ ; \*\*\*= $p \leq 0.001$ . Significance for  
 466 hydranth number at ■= $p \leq 0.05$ ; ■■= $p \leq 0.01$ ; ■■■= $p \leq 0.001$ . Significance for attachment at ▲  
 467 = $p \leq 0.05$ ; ▲▲= $p \leq 0.01$ ; ▲▲▲= $p \leq 0.001$ . Significance for feeding at ▼= $p \leq 0.05$ ; ▼▼= $p \leq 0.01$ ; ▼▼▼  
 468 = $p \leq 0.001$ .

469 Note: Error bars do not show at points where results had no variability.

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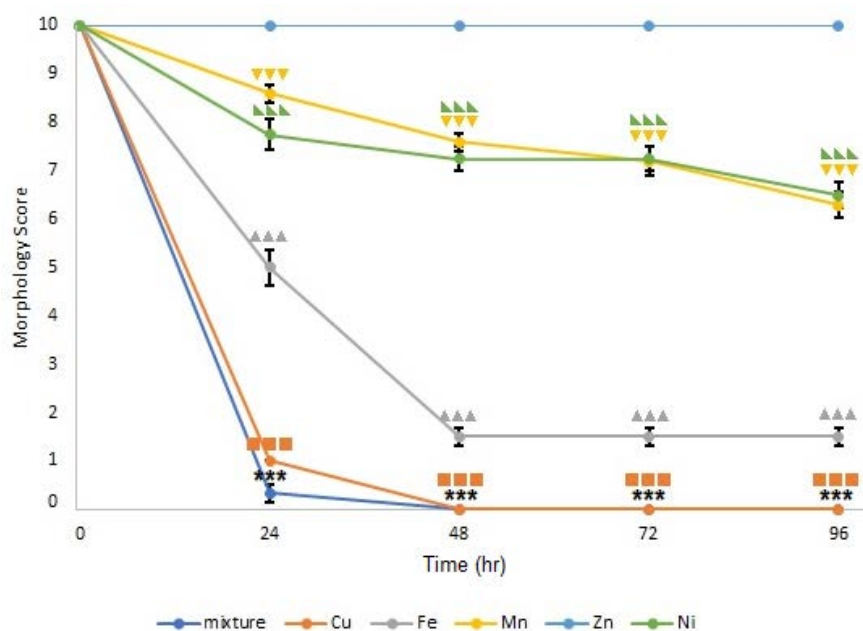
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475 Fig. 3 – Lethal and sub-lethal effects of several concentrations (0.0001x–1x) of the heavy metal  
 476 mixture found in the environment on *Hydra* morphology, hydranth number, attachment and feeding  
 477 after 96hr exposure. Points at Morphology and feeding represent the mean score (n=12) ±standard  
 478 error. Points at attachment and hydranth number represent sum (n=12). Significance for morphology  
 479 at \*= $p \leq 0.05$ ; \*\*= $p \leq 0.01$ ; \*\*\*= $p \leq 0.001$ . Significance for hydranth number at  $\square$ = $p \leq 0.05$ ;  $\blacksquare$ = $p \leq 0.01$ ;  
 480  $\blacksquare\blacksquare\blacksquare$ = $p \leq 0.001$ . Significance for attachment at  $\blacktriangle$ = $p \leq 0.05$ ;  $\blacktriangle\blacktriangle$ = $p \leq 0.01$ ;  $\blacktriangle\blacktriangle\blacktriangle$ = $p \leq 0.001$ . Significance for  
 481 feeding at  $\blacktriangledown$ = $p \leq 0.05$ ;  $\blacktriangledown\blacktriangledown$ = $p \leq 0.01$ ;  $\blacktriangledown\blacktriangledown\blacktriangledown$ = $p \leq 0.001$ .

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486 Fig. 4 – Effect of heavy metal concentrations found in the environment on *Hydra* morphology at 24, 48,  
 487 72 and 96 h exposure. Mean scores are represented the individual metals (n=8) and metal mixture  
 488 (n=12)±standard error. Significance for morphology at \*=p≤0.05; \*\*=p≤0.01; \*\*\*=p≤0.001. Significance  
 489 for Cu at ■=p≤0.05; ■■=p≤0.01; ■■■=p≤0.001. Significance for Fe at ▲=p≤0.05; ▲▲=p≤0.01; ▲▲▲  
 490 =p≤0.001. Significance for Mn at ▼=p≤0.05; ▼▼=p≤0.01; ▼▼▼=p≤0.001. Significance for Ni at ▽  
 491 =p≤0.05; ▽▽=p≤0.01; ▽▽▽=p≤0.001.

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Table 1: Estuarine and freshwater concentrations of heavy metals found in the environment.

Element	Gourock Burn (mg/l)	Kelburn Park (mg/l)	Erskine Harbour (mg/l)	Average (mg/l)
<b>Cu</b>	<LOD	0.98	0.67	0.82
<b>Fe</b>	33.78	<LOD	9.87	21.82
<b>Mn</b>	1.72	<LOD	<LOD	1.72
<b>Ni</b>	<LOD	1.74	1.42	1.58
<b>Zn</b>	<LOD	0.20	0.23	0.21

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Table 2: The concentrations of PTEs used in the exposure tests (mg/l). Based on the concentrations found in the environment.

Metal	0.001x (mg/l)	0.01x (mg/l)	0.1x (mg/l)	1x Environmental concentration (mg/l)	10x (mg/l)	100x (mg/l)	1000x (mg/l)
<b>Copper</b>	0.0005	0.005	0.05	0.5	5	-	-
<b>Iron</b>	0.003	0.03	0.3	3	30	-	-
<b>Manganese</b>	0.002	0.02	0.2	2	20	200	-
<b>Zinc</b>	-	-	0.01	0.1	1	10	100
<b>Nickel</b>	0.0005	0.005	0.05	0.5	5	50	-

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Table 3: Hydra morphology score table used to assess acute toxicity, based on the Wilby morphology score (Wilby, 1988).

10	Healthy, long tentacles and body reactive	
9	Partially contracted, slow reactions	
8	Clubbed tentacles, body slightly contracted	Alive
7	Shortened tentacles, body slightly contracted	
6	Tentacles and body shortened	
5	Totally contracted, tentacles visible	
4	Totally contracted, no visible tentacles	
3	Expanded, tentacles visible	Dead
2	Expanded, no visible tentacles	
1	Dead but intact	
0	Disintegrated	

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514 Table 4: LC<sub>50</sub>, LOEC and NOEC values based on morphology for *Hydra attenuata* exposed to heavy  
515 metals individually and as a mixture. Toxicity Threshold (TT=(NOECxLOEC)/2). Actual concentrations  
516 measured in the environment are also presented.

Metal	Concentration in environment (mg/l)	LC <sub>50</sub> (mg/l)	LOEC (mg/l)	NOEC (mg/l)	TT (mg/l)
Mixture	1x	0.045x	0.01x	0.001x	0.000005x
Copper	0.5	0.0225	0.05	0.005	0.000125
Iron	3	0.135	0.03	0.003	0.000045
Manganese	2	20	2	0.2	0.2
Zinc	0.1	31.622	10	1	5
Nickel	0.5	2.25	0.5	0.05	0.0125

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520 Table 5: Pearson correlation coefficient of heavy metal pollutants and attachment, hydranth number  
521 and feeding behaviour endpoints.

	Mixture	Cu	Fe	Mn	Ni	Zn
Attachment	-0.6742	-0.5048	-0.4882	-0.6132	-0.6666	-0.6662
Hydranth no.	-0.702	-0.5032	-0.5032	<b>-0.9257**</b>	-0.717	<b>-0.9996***</b>
Feeding	-0.7013	-0.5048	-0.5048	<b>-0.9308**</b>	-0.6848	<b>-0.922**</b>

522 Significant results indicated by bold with significance set at \*p <0.05, \*\*p <0.005, \*\*\*p <0.001



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524 Table 6: A comparison of the heavy metal concentrations found from the Clyde, Thames and Ganga  
525 rivers with the maximum acceptable limits based on EU / UK legislative requirements.

<b>Metal</b>	<b>EU / UK<sup>a</sup> (mg/l)</b>	<b>Average measured Concentration (mg/l)</b>	<b>River</b>
<b>Copper</b>	0.00376	0.5	Clyde, Glasgow, Scotland <sup>b</sup>
		0.001	Thames, London, Canada <sup>c</sup>
		0.022	Ganga, Kachlabridge, India <sup>d</sup>
<b>Iron</b>	1	3	Clyde, Glasgow, Scotland <sup>b</sup>
		0.044	Thames, London, Canada <sup>c</sup>
		0.0004	Ganga, Kachlabridge, India <sup>d</sup>
<b>Manganese</b>	-	2	Clyde, Glasgow, Scotland <sup>b</sup>
		0.011	Thames, London, Canada <sup>c</sup>
		-	Ganga, Kachlabridge, India <sup>d</sup>
<b>Zinc</b>	0.0079	0.1	Clyde, Glasgow, Scotland <sup>b</sup>
		0.002	Thames, London, Canada <sup>c</sup>
		0.00009	Ganga, Kachlabridge, India <sup>d</sup>
<b>Nickel</b>	0.0086	0.5	Clyde, Glasgow, Scotland <sup>b</sup>
		0.004	Thames, London, Canada <sup>c</sup>
		0.006	Ganga, Kachlabridge, India <sup>d</sup>

526 <sup>a</sup> SEPA (2018)

527 <sup>b</sup> Present study

528 <sup>c</sup> Environment and Engineering Services (2018)

529 <sup>d</sup> Central Water Commission (2018)

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547 Table 7: Comparison of LC<sub>50</sub> for *H. attenuata* with those from other species for selected  
 548 heavy metals.

<b>Metal</b>	<b>Organism</b>	<b>LC<sub>50</sub> (mg/l)</b>	<b>Source</b>
<b>Copper</b>	<i>Hydra attenuata</i>	0.0225	Present study
	<i>Danio rerio</i>	0.01166	Alsop & Wood (2011)
	<i>Rasbora sumatrana</i>	0.0056	Shuhaimi-Othman <i>et al.</i> , (2010)
	<i>Capoeta fusca</i>	1.1	Ebrahimpour <i>et al.</i> , (2010)
	<i>Poecilia reticulata</i>	0.0379	Shuhaimi-Othman <i>et al.</i> , (2010)
<b>Iron</b>	<i>Hydra attenuata</i>	0.135	Present study
	<i>Daphnia magna</i>	0.23	García <i>et al.</i> , (2011)
	<i>Salmo trutta</i>	0.05	Dalzell and MacFarlane (1999)
	<i>Hyalella azteca</i>	>1	Borgmann <i>et al.</i> , (2005)
<b>Manganese</b>	<i>Hydra attenuata</i>	20	Present study
	<i>Rutilus rutilus caspicus</i>	300	Hoseini <i>et al.</i> , (2014)
	<i>Mogurnda mogurnda</i>	240	Harford <i>et al.</i> , (2015)
	<i>Ceriodaphnia dubia</i>	6.2	Lasier <i>et al.</i> , (2000)
	<i>Garra gotyla gotyla</i>	3.2	Sharma & Langer (2014)
<b>Zinc</b>	<i>Hydra attenuata</i>	31.622	Present study
	<i>Danio rerio</i>	2.535	Alsop & Wood (2011)
	<i>Daphnia magna</i>	0.76	Lopes <i>et al.</i> , (2014)
	<i>Capoeta fusca</i>	13.7	Ebrahimpour <i>et al.</i> , (2010)
	<i>Lecane quadridentata</i>	0.12	Torres Guzman <i>et al.</i> , (2010)
<b>Nickel</b>	<i>Hydra attenuata</i>	2.25	Present study
	<i>Clarias gariepinus</i>	8.87	Ololade & Oginni (2010)
	<i>Hyalella azteca</i>	2	Liber <i>et al.</i> , (2011)
	<i>Danio rerio</i>	0.5898	Alsop & Wood (2011)
	<i>Chironomus dilutus</i>	119.5	Liber <i>et al.</i> , (2011)

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